# ROLE OF COX-2 IN THE CARDIOVASCULAR SYSTEM

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Prostaglandins have diverse roles in the heart and vasculature, including proinflammatory effects and vasodilation. COX-2 catalyzes the conversion of prostaglandin precursors to prostacyclin, a key vasoactive substance involved in regulation of vascular tone and perfusion. The adverse-effect profile of NSAIDs in patients with compromised cardiac function can include increased systemic vascular resistance and reductions in cardiac output, renal blood flow, and glomerular filtration (Dzau et al, 1984; Townend et al, 1995; Vane et al, 1998). In addition, prostaglandins produced by COX-2 may have a role in promoting tissue repair following injury.

#### **COX-2 EXPRESSION**

COX-2 is generally believed to be an inducible enzyme in cardiac and vascular tissues (Holmes et al, 1997; Rimarachin et al, 1994; Wong et al, 1998), although one recent in vitro study suggests that COX-2 may be constitutively expressed in rat aorta (Connolly et al, 1998). However, COX-2 knockout mice exhibit diffuse myocardial fibrosis in both ventricles and mild cardiac myofiber dropout, indicating that COX-2 deficiency may be implicated in cardiomyopathy (Dinchuk et al, 1995) and that COX-2 itself may be cardioprotective (Wu, 1998).

In a study in humans, COX-2 was not identified in hearts from healthy controls, but was identified in the myocytes and inflammatory cells of infarcted myocardial tissue in patients with heart failure secondary to sepsis or ischemic heart disease (Wong et al, 1998). In patients with heart failure secondary to dilated cardiomyopathy, COX-2 was found in fibrotic cardiac tissue, but not in myocytes or inflammatory cells.

In rabbit aorta, high-density lipoprotein (HDL) has been demonstrated to induce COX-2 expression (and increase levels of the vasodilator, prostacyclin) in smooth muscle cells (Viñals et al, 1997).

# USE OF SELECTIVE COX-2 INHIBITORS IN ANIMAL MODELS

The effects of a selective COX-2 inhibitor and a nonselective COX-1/COX-2 inhibitor (indomethacin) on vascular contractility of endothelium-denuded aorta were assessed in a murine model (Connolly et al, 1998). Both agents blocked α-agonist-induced vasoconstriction, presumably due to inhibition of COX-2. The authors speculated that COX-2 was produced constitutively in rat aorta because the experimental procedures were undertaken before the extracted tissue would have been able to induce and express the isoform.

In another preclinical study, dogs were given intravenous doses of indomethacin, 6-MNA, or a selective COX-2 inhibitor (Brooks et al, 1998). Neither 6-MNA nor indomethacin altered baseline heart rate or blood pressure, but the selective COX-2 inhibitor caused significant bradycardia (-24 bpm  $\pm$  5%) without changes in blood pressure.

# HYPOTHETICAL APPLICATION TO HUMANS OF PRECLINICAL RESULTS IN THE CARDIOVASCULAR SYSTEM

Available data on COX-2 expression in infarcts and cardiomyopathy suggest that COX-2 has a role in both inflammation and repair. Complete suppression of COX-2, as in knockout mice, may have further implications than inhibiting inflammation and may, in fact, inhibit repair. This may lead to excessive fibrosis and deleterious consequences in patients with cardiac or vascular injury.

COX-2 also appears to be involved in the control of vascular tone, participating in HDL-induced production of vasodilatory prostaglandins and  $\alpha$ -agonist-induced vasoconstriction. The consequences of inhibiting these effects remain to be determined, but could have long-term implications.

APPEARS THIS WAY ON ORIGINAL

# ROLE OF COX-2 IN THE RESPIRATORY TRACT

# ROLE OF COX-2 IN THE RESPIRATORY TRACT

The use of NSAIDs in asthmatic inflammatory conditions is controversial because inhibition of cyclooxygenase isoenzymes may increase production of leukotrienes and enhance airway responsiveness, worsening asthmatic symptoms (Szczeklik, 1983). COX-1 and COX-2 both catalyze the conversion of arachidonic acid to prostaglandin, and inhibition of either enzyme may increase leukotriene production. In addition to these effects on arachidonic acid metabolism, recent findings have demonstrated that COX-2 is involved in a variety of physiologic and regulatory processes in the pulmonary system.

## **COX-2 EXPRESSION**

Although COX-2 has a role in some noninfectious inflammatory processes in the respiratory tract (Fernández-Morata et al, 1997; Kato et al, 1998; Vadas et al, 1996), it also may have distinct regulatory functions under physiologic conditions. In animals, constitutively expressed COX-2 has been isolated from the lung (Brannon et al, 1998; Charette et al, 1995; Ermert et al, 1998). In rats, both cyclooxygenase isoforms are detected in the lungs (Ermert et al, 1998). COX-2 is noted primarily in macrophage and mast cells, smooth muscle cells of partially muscular vessels, large veins of the hilum, and bronchial epithelial cells. Extensive localization of COX-2 in the rat lung suggests that it has a physiologic, as well as an inflammatory, role in the respiratory tract. In the guinea pig, COX-2 expression is responsible for maintaining intrinsic tone in the trachea (Charette et al, 1995).

In fetal and newborn sheep lungs, COX-2 mRNA expression increased during the fetal to 1-week-old period (Brannon et al, 1998). However, COX-2 protein was not detected in the developing ovine lung. There is no evidence that COX-2 knockout mice exhibit pulmonary pathology (Dinchuk et al, 1995).

In the cultured human airway smooth muscle cell, COX-2 is not expressed under control conditions. However, COX-2 expression is stimulated by proinflammatory cytokines and bradykinin, and expression is inhibited by dexamethasone and both selective and nonselective COX-2 inhibitors (Belvisi et al, 1997; Pang and Knox, 1997a, 1997b; Vigano et al, 1997).

High levels of COX-2 mRNA are present in the normal human lung, whereas levels of COX-1 are approximately twofold lower (O'Neill and Ford-Hutchinson, 1993). In patients with stable asthma and chronic bronchitis, both cyclooxygenase isoforms are found in respiratory epithelium, but COX-2 is not upregulated (Demoly et al, 1997). In upper respiratory mucosa of patients with chronic allergic rhinitis and sinusitis and in control patients, COX-1 and COX-2 are expressed in the epithelium and neither enzyme is upregulated during this noninfectious inflammation (Demoly et al, 1998). Both cyclooxygenase isozymes are expressed in nonstimulated epithelial cells isolated from nasal polyps (Kowalski et al, 1997). Therefore, constitutive expression of COX-2 in the lungs, and its failure to increase in the presence of noninfectious inflammation, suggests a more basic, homeostatic role, rather than one restricted to supporting inflammation.

In one interesting study, lung fibroblasts from patients with idiopathic pulmonary fibrosis did not respond to inflammatory stimuli (eg, PMA, lipopolysaccharide, IL-1) with increased COX-2 expression or activity (Wilborn et al, 1995). Because one of the effects of prostaglandin  $E_2$  is to inhibit fibroblast proliferation and collagen synthesis, the authors postulated that the inability of these patients to upregulate COX-2 and increase prostaglandin production might inhibit reparative processes, allowing more opportunity for fibroblasts to lay down connective tissue and promoting the development of pulmonary fibrosis. They further suggested that "potential detrimental consequences might result from pharmacologic inhibition of COX-2, at least during the reparative phases of inflammatory injury in the lung" (Wilborn et al, 1995).

### USE OF SELECTIVE COX-2 INHIBITORS IN ANIMAL MODELS

In isolated guinea pig trachea, a selective COX-2 inhibitor and indomethacin were used to evaluate effects on intrinsic tone (Charette et al, 1995). Both agents reversed intrinsic tone, with the selective COX-2 inhibitor exhibiting greater effect, implying that COX-2 is responsible for maintaining intrinsic tone in the trachea.

# HYPOTHETICAL APPLICATION TO HUMANS OF PRECLINICAL RESULTS IN THE RESPIRATORY TRACT

Constitutive expression of COX-1 and COX-2 in the human lung clearly suggests that there is a role for these isoenzymes in maintaining normal, homeostatic functions. Furthermore, in patients with asthma or noninfectious rhinitis and sinusitis, COX-2 expression is not upregulated, as would be expected in inflammatory conditions, providing further support for a noninflammatory role for this isozyme. The consequences of inhibiting these unknown functions remain to be determined. The observation that patients with idiopathic pulmonary fibrosis are unable to induce COX-2 expression in the lung raises the question of whether selective inhibition of COX-2 might inhibit the reparative phases of inflammatory injury in the lung, or otherwise contribute to the development of fibrosis.

APPEARS THIS WAY ON ORIGINAL

# ROLE OF COX-2 IN THE PANCREAS

## ROLE OF COX-2 IN THE PANCREAS

Both COX-1 and COX-2 have been shown to be expressed in acinus cells of the pancreas in rats, both during normal conditions and during pancreatitis (Zabel-Langhennig et al, 1998). However, in pancreatic islet beta cells from hamsters, rats, and humans, COX-2 was recently shown to be constitutively expressed whereas minimal, if any, COX-1 is expressed (Sorli et al, 1998). COX-2 was also predominant during interleukin-1-stimulated conditions. The clinical relevance of these data, if any, to use of selective COX-2 inhibitors is unknown.

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# LITERATURE SEARCHES CONDUCTED TO DEVELOP THIS DOCUMENT

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The literature searches completed to develop this document were performed by Grace Johnson, PharmD, who is an Editor with Scientific Therapeutics Information, Inc, a medical communications company. Searching qualifications of Dr Johnson include training and experience received during the Doctor of Pharmacy program at the University of Michigan, an American Society of Health-System Pharmacists-accredited pharmacy practice residency completed at the University of Florida, and 3 years performing medical writing and editing as an Associate Editor or Editor with Scientific Therapeutics Information, Inc.

Online literature searches were conducted in MEDLINE (1966-1998), EMBASE (1980-1998), and BIOSIS (1997-1998), with MEDLINE being the primary reference source. All searches were limited to English language. MEDLINE search strategy included the following terms: COX-2 inhibitors, cyclooxygenase-2 inhibitors, cyclooxygenase-2 inhibitors, meloxicam, celecoxib, SC-58635, flosulide, nimesulide, SC-58125, L-748731, L-745337, SC-57666, T-614, cyclooxygenase-2 and renal, cyclooxygenase-2 and lung, cyclooxygenase-2 and inflammation, cyclooxygenase-2, COX-2, and cyclooxygenase 2 with wound healing, immune system, or hormone. EMBASE search strategy included the following terms: COX-2 inhibitor, cyclooxygenase-2 inhibitor, cyclooxygenase-2 inhibitor, cyclooxygenase-2 and renal, cyclooxygenase-2 and inflammation, cyclooxygenase-2, COX-2. BIOSIS search strategy included the following terms: cyclooxygenase-2, cyclooxygenase-2.

Additional reference sources were used when available, including references cited in publications related to this topic. Recent issues of Gastroenterology, Journal of Rheumatology, Arthritis and Rheumatism, and American Journal of Gastroenterology were reviewed for abstracts presented at the following annual meetings: Digestive Disease Week (1998), Panamerican Congress of Rheumatology

(1998), American College of Rheumatology (1997), and American College of Gastroenterology (1998), respectively.

Only selected references were used in the development of this document. Although many references were reviewed, only those cited herein are listed in the reference list at the end of the document. All cited references are available from SB upon request.

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**REFERENCES** 

#### REFERENCES

Anderson GD, Hauser SD, McGarity KL, Bremer ME, Isakson PC, Gregory SA. Selective inhibition of cyclooxygenase-2 reverses inflammation and expression of COX-2 and IL-6 in rat adjuvant arthritis. *J Clin Invest.* 1996;97:2672-2679.

Anderson RJ. Rheumatoid arthritis: Clinical features and laboratory. In: Schumacher HR, Klippel JH, Koopman WJ, eds. *Primer on the Rheumatic Diseases*. 10<sup>th</sup> ed. Atlanta, GA: The Arthritis Foundation; 1993:Chapter 10, Part B.

Asselin E, Drolet P, Fortier MA. Cellular mechanisms involved during oxytocin-induced prostaglandin  $F_{2\alpha}$  production in endometrial epithelial cells in vitro: role of cyclooxygenase-2. *Endocrinology*. 1997;138:4798-4805.

Bamba H, Ota S, Kato A, Matsuzaki F. Nonsteroidal anti-inflammatory drugs may delay the repair of gastric mucosa by suppressing prostaglandin-mediated increase of hepatocyte growth factor production. *Biochem Biophys Res Commun.* 1998:245:567-571.

Bany BM, Kennedy TG. Regulation of cyclooxygenase gene expression in rat endometrial stromal cells: the role of epidermal growth factor. *Dev Genet*. 1997;21:109-115.

Beharry K, Modanlou HD, Wold S, Rumney P, Asrat T. Cyclooxygenase (COX)-1 and COX-2 but not nitric oxide synthase (NOS) are upregulated in umbilical cord (UC) vessels in preeclamptic pregnancies (abstract). *Pediatr Res.* 1998;43:45A.

Beiche F, Scheuerer S, Brune K, Geisslinger G, Goppelt-Struebe M. Upregulation of cyclooxygenase-2 mRNA in the rat spinal cord following peripheral inflammation. *FEBS Lett.* 1996;390:165-169.

Belvisi MG, Saunders MA, Haddad EB, et al. Induction of cyclooxygenase-2 by cytokines in human cultured airway smooth muscle cells: novel inflammatory role of this cell type. *Br J Pharmacol*. 1997;120:910-916.

Bjarnason I, Sigthorsson G, Crane R, et al. COX-2 specific inhibition with MK-0966 25 or 50 mg QD does not increase intestinal permeability: a controlled study with placebo (PBO) and indomethacin 50 mg TID (INDO) (abstract). *Am J Gastroenterol*. 1998;93:1670.

Blume ED, Taylor CT, Lennon PF, Stahl GL, Colgan SP. Activated endothelial cells elicit paracrine induction of epithelial chloride secretion: 6-keto-PGF<sub>1 $\alpha$ </sub> is an epithelial secretagogue. *J Clin Invest.* 1998;102:1161-1172.

Boerboom D, Sirois J. Molecular characterization of equine prostaglandin G/H synthase-2 and regulation of its messenger ribonucleic acid in preovulatory follicles. *Endocrinology*. 1998;139:1662-1670.

Brannon TS, MacRitchie AN, Jaramillo MA, et al. Ontogeny of cyclooxygenase-1 and cyclooxygenase-2 gene expression in ovine lung. *Am J Physiol*. 1998;274:L66-L71.

Breder CD. Cyclooxygenase systems in the mammalian brain. *Ann NY Acad Sci.* 1997;813:296-301.

Breder CD, Saper CB. Expression of inducible cyclooxygenase mRNA in the mouse brain after systemic administration of bacterial lipopolysaccharide. *Brain Res.* 1996;713:64-69.

Breder CD, Dewitt D, Kraig RP. Characterization of inducible cyclooxygenase in rat brain. *J Comp Neurol*. 1995;355:296-315.

Brooks DP, Adams J, DePalma PD, Webb EF, Griswold DE, Palmer R. The selective cyclooxygenase-2 (COX-2) inhibitor, celecoxib, but not 6MNA, affects renal hemodynamic and renal function in the dog (abstract). *J Invest Med*. 1998;46:227A.

Brunel P, Hornych A, Guyene TT, Sioufi A, Turri M, Ménard J. Renal and endocrine effects of flosulide, after single and repeated administration to healthy volunteers. *Eur J Clin Pharmacol*. 1995;49:193-201.

Brzozowski T, Konturek PC, Pajdo R, Nagraba N, Szczeklik A, Konturek SJ. Role of cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2) derived prostaglandins (PG) in adaptive cytoprotection induced by mild stress (abstract). *Gastroenterology*. 1998;114:A82.

Caggiano AO, Breder CD, Kraig RP. Long-term elevation of cyclooxygenase-2, but not lipoxygenase, in regions synaptically distant from spreading depression. J Comp Neurol. 1996;376:447-462.

Cao C, Matsumura K, Yamagata K, Watanabe Y. Cyclooxygenase-2 is induced in brain blood vessels during fever evoked by peripheral or central administration of tumor necrosis factor. *Brain Res Mol Brain Res.* 1998;56:45-56.

Cao C, Matsumura K, Yamagata K, Watanabe Y. Endothelial cells of the rat brain vasculature express cyclooxygenase-2 mRNA in response to systemic interleukin-1β: a possible site of prostaglandin synthesis responsible for fever. *Brain Res.* 1996;733:263-272.

Cao C, Matsumura K, Yamagata K, Watanabe Y. Induction by lipopolysaccharide of cyclooxygenase-2 mRNA in rat brain: its possible role in the febrile response. *Brain Res.* 1995;697:187-196.

Chan TA, Morin PJ, Vogelstein B, Kinzler KW. Mechanisms underlying nonsteroidal anti-inflammatory drug-mediated apoptosis. *Proc Natl Acad Sci USA*. 1998;95:681-686.

Charette L, Misquitta C, Guay J, Riendeau D, Jones TR. Involvement of cyclooxygenase 2 (COX-2) in intrinsic tone of isolated guinea pig trachea. Can J Physiol Pharmacol. 1995;73:1561-1567.

Charpigny G, Reinaud P, Tamby JP, Créminon C, Guillomot M. Cyclooxygenase-2 unlike cyclooxygenase-1 is highly expressed in ovine embryos during the implantation period. *Biol Reprod.* 1997;57:1032-1040.

Chavez R, Bravo C, Zazueta C, et al. Ionophoretic-like properties of ketorolac for calcium. *J Pharmacol Exp Ther.* 1993;267:1134-1139.

Cohn SM, Schloemann S, Tessner T, Seibert K, Stenson WF. Crypt stem cell survival in the mouse intestinal epithelium is regulated by prostaglandins synthesized through cyclooxygenase-1. *J Clin Invest*. 1997;99:1367-1379.

Collaço-Moraes Y, Aspey B, Harrison M, de Belleroche JS. Cyclooxygenase-2 messenger RNA induction in focal cerebral ischemia. *J Cereb Blood Flow Metab*. 1996;16:1366-1372.

Connolly C, McCormick PA, Docherty JR. Effects of the selective cyclooxygenase-2 inhibitor nimesulide on vascular contractions in endothelium-denuded rat aorta. *Eur J Pharmacol.* 1998;352:53-58.

Crofford LJ. COX-1 and COX-2 tissue expression: implications and predictions. J Rheumatol. 1997;24(suppl 49):15-19.

Crofford LJ, Wilder RL, Ristimäki AP, et al. Cyclooxygenase-1 and -2 expression in rheumatoid synovial tissues: effects of interleukin-1β, phorbol ester, and corticosteroids. *J Clin Invest.* 1994;93:1095-1101.

Davies NM. Toxicity of nonsteroidal anti-inflammatory drugs in the large intestine. Dis Colon Rectum. 1995;38:1311-1321.

Demoly P, Crampette L, Lebel B, Campbell AM, Mondain M, Bousquet J. Expression of cyclooxygenases 1 and 2 proteins in upper respiratory mucosa. *Clin Exp Allergy*. 1998;28:278-283.

Demoly P, Jaffuel D, Lequeux N, et al. Prostaglandin H synthase 1 and 2 immunoreactivities in the bronchial mucosa of asthmatics. *Am J Respir Crit Care Med.* 1997;155:670-675.

DeWitt DL, Meade E, Smith WL. PGH synthase isoenzyme selectivity: the potential for safer nonsteroidal anti-inflammatory drugs. *Am J Med.* 1993;95(suppl 2A):40S-44S.

Dinchuk JE, Car BD, Focht RJ, et al. Renal abnormalities and an altered inflammatory response in mice lacking cyclooxygenase II. *Nature*. 1995;378:406-409.

Dirig DM, Konin GP, Isakson PC, Yaksh TL. Effect of spinal cyclooxygenase inhibitors in rat using the formalin test and in vitro prostaglandin  $E_2$  release. Eur J Pharmacol. 1997;331:155-160.

Donnelly MT, Hawkey CJ. Review article: COX-II inhibitors – a new generation of safer NSAIDs? *Aliment Pharmacol Ther*. 1997;11:227-236.

Dzau VJ, Packer M, Lilly LS, Swartz SL, Hollenberg NK, Williams GH. Prostaglandins in severe congestive heart failure: relation to activation of the renin-angiotensin system and hyponatremia. *N Engl J Med.* 1984;310:347-352.

Eckmann L, Stenson WF, Savidge TC, et al. Role of intestinal epithelial cells in the host secretory response to infection by invasive bacteria: bacterial entry induces epithelial prostaglandin H synthase-2 expression and prostaglandin  $E_2$  and  $F_{2\alpha}$  production. J Clin Invest. 1997;100:296-309.

Ehrlich K, Gretzer B, Respondek M, Peskar BM. Selective inhibition of cyclooxygenase 2 counteracts the protective effect of peptone in the rat stomach (abstract). *Gastroenterology*. 1997;112:A111.

Eis MJ, Watkins BM, Philip A, Welling RE. Nonsteroidal-induced benign strictures of the colon: a case report and review of the literature. *Am J Gastroenterol*. 1998;93:120-121.

Ermert L, Ermert M, Goppelt-Struebe M, et al. Cyclooxygenase isoenzyme localization and mRNA expression in rat lungs. *Am J Respir Cell Mol Biol*. 1998;18:479-488.

Evans JM, McMahon AD, Murray FE, McDevitt DG, MacDonald TM. Nonsteroidal anti-inflammatory drugs are associated with emergency admission to hospital for colitis due to inflammatory bowel disease. *Gut.* 1997;40:619-622.

Fernández-Morata JC, Mullol J, Juan M, et al. Gene expression of COX-1 and COX-2 in human nasal mucosa: modulation by cytokines and steroids (abstract).  $Eur\ Respir\ J.\ 1997;10(suppl):444S.$ 

Ferraz JG, Sharkey KA, Reuter BK, et al. Induction of cyclooxygenase 1 and 2 in the rat stomach during endotoxemia: role in resistance to damage. Gastroenterology. 1997;113:195-204.

Fuchs AR, Rust W, Fields MJ. Cyclooxygenase-1 and -2 transcript accumulation in bovine uterine tissues during the second half of pregnancy and parturition (abstract). *Biol Reprod.* 1998;58(suppl 1):122.

Fuentes A, Spaziani EP, O'Brien WF. The expression of cyclooxygenase-2 (COX-2) in amnion and decidua following spontaneous labor. *Prostaglandins*. 1996;52:261-267.

Gibb W, Sun M. Localization of prostaglandin H synthase type 2 protein and mRNA in term human fetal membranes and decidua. *J Endocrinol*. 1996;150:497-503.

Gibson GR, Whitacre EB, Ricotti CA. Colitis induced by nonsteroidal anti-inflammatory drugs: report of four cases and review of the literature. *Arch Intern Med.* 1992;152:625-632.

Gilroy DW, Tomlinson A, Willoughby DA. Differential effects of inhibition of isoforms of cyclooxygenase (COX-1, COX-2) in chronic inflammation. *Inflamm Res.* 1998;47:79-85.

Gretzer B, Ehrlich K, Maricic N, Lambrecht N, Respondek M, Peskar BM. Selective cyclooxygenase-2 inhibitors and their influence on the protective effect of a mild irritant in the rat stomach. *Br J Pharmacol*. 1998a;123:927-935.

Gretzer B, Knorth H, Chantrain M, et al. Effects of diclofenac and L-745,337, a selective cyclooxygenase-2 inhibitor, on prostaglandin E<sub>2</sub> formation in tissue from human colonic mucosa and chronic bursitis (abstract). *Gastroenterology*. 1998b;114:A139.

Guan Y, Chang M, Cho W, et al. Cloning, expression, and regulation of rabbit cyclooxygenase-2 in renal medullary interstitial cells. *Am J Physiol.* 1997;273:F18-F26.

Harding P, Sigmon DH, Alfie ME, et al. Cyclooxygenase-2 mediates increased renal renin content induced by low-sodium diet. *Hypertension*. 1997;29(part 2):297-302.

Harris RC, McKanna JA, Akai Y, Jacobson HR, Dubois RN, Breyer MD. Cyclooxygenase-2 is associated with the macula densa of rat kidney and increases with salt restriction. *J Clin Invest.* 1994;94:2504-2510.

Hartner A, Goppelt-Struebe M, Hilgers KF. Coordinate expression of cyclooxygenase-2 and renin in the rat kidney in renovascular hypertension. *Hypertension*. 1998;31(part 2):201-205.

Hawkey CJ, Karrasch JA, Szczepanski L, et al. Omeprazole compared with misoprostol for ulcers associated with nonsteroidal anti-inflammatory drugs. Omeprazole versus Misoprostol for NSAID-induced Ulcer Management (OMNIUM) Study Group. *N Engl J Med.* 1998;338:727-734.

Hay CH, Trevethick MA, Wheeldon A, Bowers JS, de Belleroche JS. The potential role of spinal cord cyclooxygenase-2 in the development of Freund's complete adjuvant-induced changes in hyperalgesia and allodynia. *Neuroscience*. 1997;78:843-850.

Hayaishi O. Molecular mechanism of sleep-wake regulation: roles of prostaglandins  $D_2$  and  $E_2$ . FASEB J. 1991;5:2575-2581.

Heerdink ER, Leufkens HG, Herings RMC, Ottervanger JP, Stricker BHC, Bakker A. NSAIDs associated with increased risk of congestive heart failure in elderly patients taking diuretics. *Arch Intern Med.* 1998;158:1108-1112.

Hendel J, Nielsen OH. Expression of cyclooxygenase-2 mRNA in active inflammatory bowel disease. Am J Gastroenterol. 1997;92:1170-1173.

Holmes DR, Wester W, Thompson RW, Reilly JM. Prostaglandin  $E_2$  synthesis and cyclooxygenase expression in abdominal aortic aneurysms. *J Vasc Surg*. 1997;25:810-815.

Holtz ML, Kindy MS, Craddock S, Moore RW, Pettigrew LC. Induction of PGH synthase and c-fos mRNA during early reperfusion of ischemic rat brain. *Mol Brain Res.* 1996;35:339-343.

Houston MC, Weir M, Gray J, et al. The effects of nonsteroidal anti-inflammatory drugs on blood pressures of patients with hypertension controlled by verapamil. *Arch Intern Med.* 1995;155:1049-1054.

Hunt R, Bowen B, James C, et al. COX-2 specific inhibition with MK-0966 25 or 50 mg QD over 4 weeks does not increase fecal blood loss: a controlled study with placebo and ibuprofen 800 mg TID (abstract). *Am J Gastroenterol*. 1998;93:1671.

Iadecola C, Ross ME. Role of iNOS and COX-2 gene expression in ischemic brain injury (abstract). *J Neurochem*. 1998;70(suppl 1):S17.

Ichitani Y, Shi T, Haeggstrom JZ, Samuelsson B, Hökfelt T. Increased levels of cyclooxygenase-2 mRNA in the rat spinal cord after peripheral inflammation: an in situ hybridization study. *Neuroreport*. 1997;8:2949-2952.

Iñiguez MA, Pablos JL, Carreira PE, Cabré F, Gomez-Reino JJ. Detection of COX-1 and COX-2 isoforms in synovial fluid cells from inflammatory joint diseases. *Br J Rheumatol.* 1998;37:773-778.

Insel PA. Analgesic-antipyretics and anti-inflammatory agents: drugs employed in the treatment of rheumatoid arthritis and gout. In: Goodman Gilman A, Rall TW, Nies AS, Taylor P, eds. Goodman and Gilman's The Pharmacological Basis of Therapeutics. 8th ed. Elmsford, NY: Pergamon Press, Inc; 1990:638-681.

Johnson AG, Simons LA, Simons J, Friedlander Y, McCallum J. Nonsteroidal antiinflammatory drugs and hypertension in the elderly: a community-based crosssectional study. *Br J Clin Pharmacol*. 1993;35:455-459.

Jouzeau JY, Terlain B, Abid A, Nédélec E, Netter P. Cyclooxygenase isoenzymes: how recent findings affect thinking about nonsteroidal anti-inflammatory drugs. *Drugs.* 1997;53:563-582.

Kaplan B, Swain RA. NSAIDs: are there any differences? Arch Fam Med. 1993;2:1167-1174.

Kargman S, Charleson S, Cartwright M, et al. Characterization of prostaglandin G/H synthase 1 and 2 in rat, dog, monkey, and human gastrointestinal tracts. *Gastroenterology*. 1996;111:445-454.

Kato T, Ishiguro N, Iwata H, Kojima T, Ito T, Naruse K. Upregulation of COX-2 expression by uni-axial cyclic stretch in human lung fibroblast cells. *Biochem Biophys Res Commun.* 1998;244:615-619.

Kaufman HL, Fischer AH, Carroll M, Becker JM. Colonic ulceration associated with nonsteroidal anti-inflammatory drugs: report of three cases. *Dis Colon Rectum.* 1996;39:705-710.

Kaufmann WE, Andreasson KI, Isakson PC, Worley PF. Cyclooxygenases and the central nervous system. *Prostaglandins*. 1997;54:601-624.

Kaufmann WE, Worley PF, Pegg J, Bremer M, Isakson P. COX-2, a synaptically induced enzyme, is expressed by excitatory neurons at postsynaptic sites in rat cerebral cortex. *Proc Natl Acad Sci USA*. 1996;93:2317-2321.

Kishimoto Y, Wada K, Nakamoto K, Kawasaki H, Hasegawa J. Levels of cyclooxygenase-1 and -2 mRNA expression at various stages of acute gastric injury induced by ischemia-reperfusion in rats. *Arch Biochem Biophys.* 1998;352:153-157.

Kishimoto Y, Wada K, Nakamoto K, et al. Quantitative analysis of cyclooxygenase-2 gene expression on acute gastric injury induced by ischemia-reperfusion in rats. *Life Sci.* 1997;60:127-133.

Klein T, Ullrich V, Pfeilschifter J, Nüsing R. On the induction of cyclooxygenase-2, inducible nitric oxide synthase and soluble phospholipase  $A_2$  in rat mesangial cells by a nonsteroidal anti-inflammatory drug: the role of cyclic AMP. *Mol Pharmacol*. 1998;53:385-391.

Kömhoff M, Gröne H-J, Klein T, Seyberth HW, Nüsing RM. Localization of cyclooxygenase-1 and -2 in adult and fetal human kidney: implication for renal function. *Am J Physiol*. 1997;272:F460-F468.

Kowalski ML, Kowalewski R, Pawliczak R, Woszczek J, Grzegorczyk J. Human nasal epithelial cells cultured from nasal polyps express both cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2) mRNA (abstract). *Eur Respir J*. 1997;10(suppl):255S.

Lancaster-Smith MJ, Jaderberg ME, Jackson DA. Ranitidine in the treatment of nonsteroidal anti-inflammatory drug associated gastric and duodenal ulcers. *Gut*. 1991;32:252-255.

Langenbach R, Morham SG, Tiano HF, et al. Prostaglandin synthase 1 gene disruption in mice reduces arachidonic acid-induced inflammation and indomethacin-induced gastric ulceration. *Cell.* 1995;83:483-492.

Lanza F, Simon T, Quan H, et al. Selective inhibition of cyclooxygenase-2 (COX-2) with MK-0966 (250 mg QD) is associated with less gastroduodenal damage than aspirin (ASA) 650 mg QID or ibuprofen (IBU) 800 mg TID (abstract). Gastroenterology. 1997a;112:A194.

Lanza FL, Rack MF, Callison DA, et al. A pilot endoscopic study of the gastroduodenal effects of SC-58635, a novel COX-2-selective inhibitor (abstract). *Gastroenterology*. 1997b;112:A194.

Li DY, Hardy P, Abran D, et al. Key role for cyclooxygenase-2 in  $PGE_2$  and  $PGF_{2\alpha}$  receptor regulation and cerebral blood flow of the newborn. *Am J Physiol*. 1997;273:R1283-R1290.

Lichtenberger LM, Wang ZM, Romero JJ, et al. Non-steroidal anti-inflammatory drugs (NSAIDs) associate with zwitterionic phospholipids: insight into the mechanism and reversal of NSAID-induced gastrointestinal injury. *Nat Med*. 1995;1:602-603.

Lim H, Paria BC, Das SK, et al. Multiple female reproductive failures in cyclooxygenase 2-deficient mice. *Cell.* 1997;91:197-208.

Liu J, Carrière PD, Doré M, Sirois J. Prostaglandin G/H synthase-2 is expressed in bovine preovulatory follicles after the endogenous surge of luteinizing hormone. *Biol Reprod.* 1997;57:1524-1531.

Lugea A, Antolin M, Mourelle M, Guarner F, Malagelada JR. Deranged hydrophobic barrier of the rat gastroduodenal mucosa after parenteral nonsteroidal anti-inflammatory drugs. *Gastroenterology*. 1997;112:1931-1939.

Macchia L, Di Paola R, Guerrese MC, et al. Expression of prostaglandin endoperoxide H synthase 1 and 2 in human placenta at term. *Biochem Biophys Res Commun.* 1997;233:496-501.

Mahida YR, Beltinger J, Makh S, et al. Adult human colonic subepithelial myofibroblasts express extracellular matrix proteins and cyclooxygenase-1 and -2. *Am J Physiol.* 1997;273:G1341-G1348.

Mahmud T, Scott DL, Bjarnason I. A unifying hypothesis for the mechanism of NSAID related gastrointestinal toxicity. *Ann Rheum Dis.* 1996;55:211-213.

Majerus PW. Prostaglandins: critical roles in pregnancy and colon cancer. *Curr Biol.* 1998;8:R87-R89.

Majima M, Isono M, Ikeda Y, et al. Significant roles of inducible cyclooxygenase (COX)-2 in angiogenesis in rat sponge implants. *Jpn J Pharmacol.* 1997;75:105-114.

Marcheselli VL, Bazan NG. Sustained induction of prostaglandin endoperoxide synthase-2 by seizures in hippocampus. *J Biol Chem.* 1996;271:24794-24799.

Maricic N, Ehrlich K, Schuligoi R, Respondek M, Peskar BM. Cyclooxygenase-2-derived prostaglandins contribute to mucosal resistance to ischemia/reperfusion injury in the rat stomach (abstract). *Gastroenterology*. 1998;114:A215.

Marini U, Spotti D. Gastric tolerability of nimesulide: a double-blind comparison of 2 oral dosage regimens and placebo. *Drugs.* 1993;46(suppl 1):249-252.

Matsumura K, Cao C, Watanabe Y. Possible role of cyclooxygenase-2 in the brain vasculature in febrile response. *Ann NY Acad Sci.* 1997;813:302-306.

McCown TJ, Knapp DJ, Crews FT. Inferior collicular seizure generalization produces site-selective cortical induction of cyclooxygenase-2 (COX-2). *Brain Res.* 1997;767:370-374.

McKanna JA, Zhange MZ, Wang JL, Cheng HF, Harris RC. Constitutive expression of cyclooxygenase-2 in rat vas deferens. *Am J Physiol.* 1998;275:R227-R233.

Mizuno H, Sakamoto C, Matsuda K, et al. Induction of cyclooxygenase 2 in gastric mucosal lesions and its inhibition by the specific antagonist delays healing in mice. *Gastroenterology*. 1997;112:387-397.

Morham SG, Langenbach R, Loftin CD, et al. Prostaglandin synthase 2 gene disruption causes severe renal pathology in the mouse. *Cell.* 1995;83:473-482.

Morteau O, Morham S, Sellon R, Smithies O, Sartor RB. Absence of spontaneous gastrointestinal inflammation in cyclooxygenase-1 and in cyclooxygenase-2 deficient mice (abstract). *Gastroenterology*. 1997a;112:A1045.

Morteau O, Morham S, Sellon R, Boroviov J, Smithies O, Sartor RB. Genetic deficiency in cyclooxygenase-2 but not in cyclooxygenase-1 exacerbates DSS-induced acute colitis in mice (abstract). *Gastroenterology*. 1997b;112:A1046.

Nogawa S, Zhang F, Ross ME, Iadecola C. Cyclooxygenase-2 gene expression in neurons contributes to ischemic brain damage. *J Neurosci*. 1997;17:2746-2755.

Norwood VF. Progressive postnatal renal dysplasia in cyclooxygenase-2 deficient mice (abstract 1828). *Pediatr Res.* 1998;43(4 Part 2):312A.

O'Neill GP, Ford-Hutchinson AW. Expression of mRNA for cyclooxygenase-1 and cyclooxygenase-2 in human tissues. *FEBS Lett.* 1993;330:156-160.

Once Y, Miyaura C, Kaminakayashiki T, et al. IL-13 and IL-4 inhibit bone resorption by suppressing cyclooxygenase-2-dependent prostaglandin synthesis in osteoblasts. *J Immunol*. 1996;156:758-764.

Öst M, Uhl E, Carlsson M, Gidlöf A, Söderkvist P, Sirsjö A. Expression of mRNA for phospholipase A<sub>2</sub>, cyclooxygenases, and lipoxygenases in cultured human umbilical vascular endothelial and smooth muscle cells and in biopsies from umbilical arteries and veins. *J Vasc Res.* 1998;35:150-155.

Osuka K, Suzuki Y, Watanabe Y, Takayasu M, Yoshida J. Inducible cyclooxygenase expression in canine basilar artery after experimental subarachnoid hemorrhage. *Stroke.* 1998;29:1219-1222.

Pang L, Knox AJ. Effect of interleukin- $1\beta$ , tumour necrosis factor- $\alpha$  and interferon- $\gamma$  on the induction of cyclooxygenase-2 in cultured human airway smooth muscle cells. *Br J Pharmacol*. 1997a;121:579-587.

Pang L, Knox AJ. PGE<sub>2</sub> release by bradykinin in human airway smooth muscle cells: involvement of cyclooxygenase-2 induction. *Am J Physiol*. 1997b;273:L1132-L1140.

Parfenova H, Eidson TH, Leffler CW. Upregulation of COX-2 in cerebral microvascular endothelial cells by smooth muscle cell signals. *Am J Physiol*. 1997;273:C277-C288.

Park JM, Yang T, Arend LJ, Smart AM, Schnermann JB, Briggs JP. Cyclooxygenase-2 is expressed in bladder during fetal development and stimulated by outlet obstruction. *Am J Physiol*. 1997;273:F538-F544.

Peri KG, Hardy P, Li DY, Varma DR, Chemtob S. Prostaglandin G/H synthase-2 is a major contributor of brain prostaglandins in the newborn. *J Biol Chem*. 1995;270:24615-24620.

Piazza GA, Alberts DS, Hixson LJ, et al. Sulindac sulfone inhibits azoxymethane-induced colon carcinogenesis in rats without reducing prostaglandin levels. *Cancer Res.* 1997;57:2909-2915.

Pope JE, Anderson JJ, Felson DT. A meta-analysis of the effects of nonsteroidal anti-inflammatory drugs on blood pressure. *Arch Intern Med.* 1993;153:477-484.

Resnick DK, Graham SH, Dixon CE, Marion DW. The role of cyclooxygenase 2 in acute spinal cord injury (abstract). *J Neurotrauma*. 1998;15:20.

Reuter BK, Asfaha S, Buret A, Sharkey KA, Wallace JL. Exacerbation of inflammation-associated colonic injury in rat through inhibition of cyclooxygenase-2. *J Clin Invest.* 1996;98:2076-2085.

Richards JS. Editorial: sounding the alarm – does induction of prostaglandin endoperoxide synthase-2 control the mammalian ovulatory clock? *Endocrinology*. 1997;138:4047-4048.

Rimarachin JA, Jacobson JA, Szabo P, Maclouf J, Creminon C, Weksler BB. Regulation of cyclooxygenase-2 expression in aortic smooth muscle cells. *Arterioscler Thromb.* 1994;14:1021-1031.

Rossat J, Maillard M, Nussberger J, Drower E, Brunner H-R, Burnier M. Acute renal effects of selective inhibition of cyclooxygenase-2 in healthy salt-depleted subjects (abstract). *J Am Soc Nephrol*. 1998;9:346A.

Sairanen T, Ristimäki A, Karjalainen-Lindsberg ML, Paetau A, Kaste M, Lindsberg PJ. Cyclooxygenase-2 is induced globally in infarcted human brain. *Ann Neurol*. 1998;43:738-747.

San Miguel P, Laudanno OM, Guastalli G, Esnarriaga JM, Cesolari JA. Anti-inflammatory drugs: COX-2 selective and stress-induced gastric lesions (abstract). *Gastroenterology*. 1998;114:A276.

Sano H, Hla T, Maier JA, et al. In vivo cyclooxygenase expression in synovial tissues of patients with rheumatoid arthritis and osteoarthritis and rats with adjuvant and streptococcal cell wall arthritis. *J Clin Invest.* 1992;89:97-108.

Sarrazin P, de Brum-Fernandes AJ. Isolation of mature human osteoclasts and characterization of cyclooxygenase isoforms expression (abstract). *Bone*. 1998;23(suppl):S338.

Sasaki E, Pai R, Komurasaki T, et al. Induction of cyclooxygenase-2 in rat gastric epithelial cells by EGF, epiregulin and bFGF (abstract). *Gastroenterology*. 1998;114:A277.

Sato T, Kirimura Y, Mori Y. The co-culture of dermal fibroblasts with human epidermal keratinocytes induces increased prostaglandin E<sub>2</sub> production and cyclooxygenase-2 activity in fibroblasts. *J Invest Dermatol.* 1997;109:334-339.

Sawaoka H, Tsuji S, Tsujii M, et al. Expression of the cyclooxygenase-2 gene in gastric epithelium. *J Clin Gastroenterol*. 1997;25(suppl 1):S105-S110.

Schmassmann A. Mechanisms of ulcer healing and effects of nonsteroidal anti-inflammatory drugs. *Am J Med*. 1998;104(suppl 3A):43S-51S.

Schmassmann A, Peskar BM, Stettler C, et al. Effects of inhibition of prostaglandin endoperoxide synthase-2 in chronic gastrointestinal ulcer models in rats.  $Br\ J$  Pharmacol. 1998;123:795-804.

Schmassmann A, Tarnawski A, Peskar BM, Varga L, Flogerzi B, Halter H. Influence of acid and angiogenesis on kinetics of gastric ulcer healing in rats: interaction with indomethacin. *Am J Physiol*. 1995;268:G276-G285.

Schneider A, Stahl RAK. Cyclooxygenase-2 (COX-2) and the kidney: current status and potential perspectives. *Nephrol Dial Transplant*. 1998;13:10-12.

Seibert K, Zhang Y, Leahy K, et al. Pharmacological and biochemical demonstration of the role of cyclooxygenase 2 in inflammation and pain. *Proc Natl Acad Sci USA*. 1994;91:12013-12017.

Shah AA, Murray FE, Thjodleifsson B, et al. Nimesulide, a selective COX-2 inhibitor, causes less gastrointestinal damage compared with naproxen: a prospective study in human volunteers (abstract). *Gastroenterology*. 1998;114:A283.

Shigeta J, Takahashi S, Ishikawa M, Okabe S. Role of cyclooygenase-2 (COX-2) in the healing of gastric ulcers in rats (abstract). *Gastroenterology*. 1998;114:A285.

Singer II, Kawka DW, Schloemann S, Tessner T, Riehl T, Stenson WF. Cyclooxygenase 2 is induced in colonic epithelial cells in inflammatory bowel disease. *Gastroenterology*. 1998;115:297-306.

Sirois J. Induction of prostaglandin endoperoxide synthase-2 by human chorionic gonadotropin in bovine preovulatory follicles in vivo. *Endocrinology*. 1994;135:841-848.

Sirois J, Doré M. The late induction of prostaglandin G/H synthase-2 in equine preovulatory follicles supports its role as a determinant of the ovulatory process. *Endocrinology*. 1997;138:4427-4434.

Sirois J, Richards JS. Purification and characterization of a novel, distinct isoform of prostaglandin endoperoxide synthase induced by human chorionic gonadotropin in granulosa cells of rat preovulatory follicles. *J Biol Chem.* 1992;267:6382-6388.

Slater DM, Allport V, Bennett P. Changes in the expression of the type-2 but not the type-1 cyclooxygenase enzyme in chorion-decidua with the onset of labour. Br J Obstet Gynaecol. 1998;105:745-748.

Slater DM, Berger LC, Newton R, Moore GE, Bennett PR. Expression of cyclooxygenase types 1 and 2 in human fetal membranes at term. *Am J Obstet Gynecol.* 1995;172:77-82.

Slater DM, Berger LC, Moore GE, Bennett PR. Implication of mRNA binding proteins in the regulation of cyclooxygenase in human amnion at term. *Biochem Biophys Res Commun.* 1994;203:67-71.

Smith G, Roberts R, Hall C, Nuki G. Reversible ovulatory failure associated with the development of luteinized unruptured follicles in women with inflammatory arthritis taking nonsteroidal anti-inflammatory drugs. *Br J Rheumatol*. 1996;35:458-462.

Smith WL, DeWitt DL. Biochemistry of prostaglandin endoperoxide H synthase-1 and synthase-2 and their differential susceptibility to nonsteroidal anti-inflammatory drugs. *Semin Nephrol.* 1995;15:179-194.

Somasundaram S, Rafi S, Hayllar J, et al. Mitochondrial damage: a possible mechanism of the "topical" phase of NSAID-induced injury to rat intestine. *Gut*. 1997;41:344-353.

APPEARS THIS WAY ON ORIGINAL

Song JH, Sirois J, Houde A, Murphy BD. Cloning, developmental expression, and immunohistochemistry of cyclooxygenase 2 in the endometrium during embryo implantation and gestation in the mink (*Mustela vison*). *Endocrinology*. 1998;139:3629-3636.

Sorli CH, Zhang HJ, Armstrong MB, Rajotte RV, Maclouf J, Robertson RP. Basal expression of cyclooxygenase-2 and nuclear factor-interleukin 6 are dominant and coordinately regulated by interleukin 1 in the pancreatic islet. *Proc Natl Acad Sci USA*. 1998;95:1788-1793.

Stamm C, Burkhalter E, Pearce W, et al. Benign colonic ulcers associated with nonsteroidal anti-inflammatory drug ingestion. *Am J Gastroenterol*. 1994;89:2230-2233.

Steinhäuslin F, Munafo A, Buclin T, Macciocchi A, Biollaz J. Renal effects of nimesulide in furosemide-treated subjects. *Drugs*. 1993;46(suppl 1):257-262.

Stenson WF. Cyclooxygenase 2 and wound healing in the stomach (editorial). *Gastroenterology*. 1997;112:645-648.

Swaisgood CM, Zu HX, Perkins DJ, et al. Coordinate expression of inducible nitric oxide synthase and cyclooxygenase-2 genes in uterine tissues of endotoxin-treated pregnant mice. *Am J Obstet Gynecol.* 1997;177:1253-1262.

Szczeklik A. Anti-cyclooxygenase agents and asthma. J Asthma. 1983;20:23-29.

Tarnawski A, Kidao J, Jaafar S, et al. Expression and localization of cyclooxygenase-2 in normal and ulcerated human gastric mucosa, and in cultured human endothelial cells (abstract). *Gastroenterology*. 1997;112:A309.

Tocco G, Freire-Moar J, Schreiber SS, Sakhi SH, Aisen PS, Pasinetti GM. Maturational regulation and regional induction of cyclooxygenase-2 in rat brain: implications for Alzheimer's disease. *Exp Neurol*. 1997;144:339-349.

Townend JN, Doran J, Lote CJ, Davies MK. Peripheral haemodynamic effects of inhibition of prostaglandin synthesis in congestive heart failure and interactions with captopril. *Br Heart J.* 1995;73:434-441.

Tsuji M, DuBois RN. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. *Cell.* 1995;83:493-501.

Tsuji S, Sun WH, Sawaoka H, et al. A specific inhibitor for cyclooxygenase-2 delays gastric ulcer healing (abstract). *Gastroenterology*. 1997;112:A316.

Twomey BM, Dale MM. Cyclooxygenase-independent effects of nonsteroidal antiinflammatory drugs on the neutrophil respiratory burst. *Biochem Pharmacol*. 1992;43:413-418.

Ukawa H, Hirata T, Kobayashi H, Yamakuni H, Takeuchi K. Effects of COX-2-selective and NO-releasing NSAIDs on mucosal ulcerogenic and healing responses in rat stomachs (abstract). *Gastroenterology*. 1997;112:A317.

Vadas P, Stefanski E, Wloch M, Grouix B, Van Den Bosch H, Kennedy B. Secretory non-pancreatic phospholipase A<sub>2</sub> and cyclooxygenase-2 expression by tracheobronchial smooth muscle cells. *Eur J Biochem*. 1996;235:557-563.

Vane JR, Bakhle YS, Botting RM. Cyclooxygenases 1 and 2. Annu Rev Pharmacol Toxicol. 1998;38:97-120.

Viganò T, Habib A, Hernandez A, et al. Cyclooxygenase-2 and synthesis of PGE<sub>2</sub> in human bronchial smooth-muscle cells. *Am J Respir Crit Care Med.* 1997;155:864-868.

Viñals M, Martínez-González J, Badimon JJ, Badimon L. HDL-induced prostacyclin release in smooth muscle cells is dependent on cyclooxygenase-2 (COX-2). Arterioscler Thromb Vasc Biol. 1997;17:3481-3488.

Von Knethen A, Brüne B. Cyclooxygenase-2: an essential regulator of Nomediated apoptosis. *FASEB J.* 1997;11:887-895.

Walan A, Bader J-P, Classen M, et al. Effect of omeprazole and ranitidine on ulcer healing and relapse rates in patients with benign gastric ulcer. *N Engl J Med*. 1989;320:69-75.

Wallace JL. Nonsteroidal anti-inflammatory drugs and gastroenteropathy: the second hundred years. *Gastroenterology*. 1997;112:1000-1016.

Wallace JL, Bak A, McKnight W, Asfaha S, Sharkey KA, MacNaughton WK. Cyclooxygenase 1 contributes to inflammatory responses in rats and mice: implications for gastrointestinal toxicity. *Gastroenterology*. 1998;115:101-109.

Wetzka B, Nüsing R, Charnock-Jones DS, Schäfer W, Zahradnik HP, Smith SK. Cyclooxygenase-1 and -2 in human placenta and placental bed after normal and preeclamptic pregnancies. *Hum Reprod.* 1997;12:2313-2320.

Whelton A, Hamilton CW. Nonsteroidal anti-inflammatory drugs: effects on kidney function. *J Clin Pharmacol*. 1991;31:588-598.

Wilborn J, Crofford LJ, Burdick MD, Kunkel SL, Strieter RM, Peters-Golden M. Cultured lung fibroblasts isolated from patients with idiopathic pulmonary fibrosis have a diminished capacity to synthesize prostaglandin E<sub>2</sub> and to express cyclooxygenase-2. *J Clin Invest.* 1995;95:1861-1868.

Willingale HL, Gardiner NJ, McLymont N, Giblett S, Grubb BD. Prostanoids synthesized by cyclooxygenase isoforms in rat spinal cord and their contribution to the development of neuronal hyperexcitability. *Br J Pharmacol*. 1997;122:1593-1604.

Wong SCY, Fukuchi M, Melnyk P, Rodger I, Giaid A. Induction of cyclooxygenase-2 and activation of nuclear factor-kB in myocardium of patients with congestive heart failure. *Circulation*. 1998;98:100-103.

Wu KK. Cyclooxygenase-2 induction in congestive heart failure: friend or foe? *Circulation*. 1998;98:95-96.

Xiao CW, Murphy BD, Sirois J, Goff AK. Downregulation of oxytocin receptor and oxytocin-induced prostaglandin secretion, COX-2 and prostaglandin F synthase expression by interferon-τ bovine endometrial cells in vitro (abstract). *Biol Reprod.* 1998;58(suppl 1):174.

Yamagata K, Andreasson KI, Kaufmann WE, Barnes CA, Worley PF. Expression of a mitogen-inducible cyclooxygenase in brain neurons: regulation by synaptic activity and glucocorticoids. *Neuron*. 1993;11:371-386.

Yamamoto T, Nozaki-Taguchi N. Analysis of the effects of cyclooxygenase (COX)-1 and COX-2 in spinal nociceptive transmission using indomethacin, a nonselective COX inhibitor, and NS-398, a COX-2 selective inhibitor. *Brain Res.* 1996;739:104-110.

Yang T, Singh I, Pham H, et al. Regulation of cyclooxygenase expression in the kidney by dietary salt intake. Am J Physiol. 1998;274:F481-F489.

Yeomans ND, Cook GA, Giraud AS. Selective COX-2 inhibitors: are they safe for the stomach? (editorial). *Gastroenterology*. 1998;115:227-229.

Zabel-Langhennig A, Holler B, Engeland K, Mössner J. Regulation of transcription of the two cyclooxygenase isoforms in pancreas acini (abstract). *Gastroenterology*. 1998;114:A512.

Zhang M-Z, Wang J-L, Cheng H-F, Harris RC, McKanna JA. Cyclooxygenase-2 in rat nephron development. *Am J Physiol.* 1997;273:F994-F1002.

Zuo J, Lei ZM, Rao CV, Pietrantoni M, Cook VD. Differential cyclooxygenase-1 and -2 gene expression in human myometria from preterm and term deliveries. *J Clin Endocrinol Metab.* 1994;79:894-899.

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#### **EXECUTIVE SUMMARY**

Celecoxib (tradename Celebrex<sup>TM</sup>) is an oral anti-inflammatory and analgesic product intended for use in the treatment of the signs and symptoms of osteoarthritis (OA) and rheumatoid arthritis (RA) and for the management of pain. It was developed to provide anti-inflammatory and analgesic effects comparable to commonly used non-steroidal anti-inflammatory agents drugs (NSAIDs) but without the gastrointestinal and platelet effects of NSAIDs. The clinical development program for celecoxib included more than 13,000 patients and healthy volunteers enrolled in over 50 different studies. The overall results of this clinical program demonstrated that celecoxib is comparable in efficacy to naproxen and is superior to naproxen, diclofenac and ibuprofen in the occurrence of upper gastrointestinal (UGI) ulceration, ulcer complications and effects on platelets. Further, celecoxib was not found to be associated with increases in other clinically significant adverse reactions compared to NSAIDs. Thus, celecoxib has been shown to provide a significant therapeutic benefit over commonly prescribed NSAIDs used in the treatment of OA, RA and pain.

Celecoxib is a highly selective inhibitor of cyclooxygenase-2 (COX-2), one of two known isoforms of cyclooxygenase: COX-1 and COX-2. Commonly used NSAIDs such as aspirin, naproxen and ibuprofen are non-selective inhibitors of COX-1 and COX-2.

(1) They are effective anti-arthritic agents but produce clinically-limiting side effects due to their inhibition of COX-1 at therapeutic doses. (2-4)

The clinical program examined the human pharmacokinetics, efficacy, and safety profile of celecoxib with a focus on its effects on the GI tract, platelets and kidney.

Celecoxib is a low solubility, high permeability drug that is rapidly absorbed, has a large volume of distribution (consistent with high tissue uptake) and is extensively metabolized by the liver to inactive metabolites that are excreted primarily in the feces and to a lesser extent in the urine. Pharmacokinetics are linear; the elimination half life is about 10 hours and steady state levels are reached by five days with the therapeutic dose and regimen. The pharmacokinetics in subgroup populations such as the elderly were studied and well described. No clinically significant drug-drug interactions were observed with compounds commonly used by the intended patient population, including warfarin and methotrexate.

Five pivotal trials of 6 or 12 weeks duration uniformly showed that celecoxib was superior to placebo in treating OA of the knee and/or hip and similar to a full therapeutic dose of naproxen. Maximal efficacy was obtained with 200 mg per day administered as a single dose or divided doses. Replicate 12-week studies showed that celecoxib at 100 mg or 200 mg BID was superior to placebo in treating RA and similar to naproxen. A 6-month trial confirmed the durability of response with celecoxib. The data from four

trials using an acute post-surgery pain model and the chronic pain data from the aforementioned arthritis trials collectively demonstrated that celecoxib has analgesic properties. The post-oral surgery studies showed that celecoxib had an onset of action within one hour and significant relief of OA pain was demonstrated within 24 hours after the first dose.

Over 4,500 patients with OA or RA participated in endoscopy trials designed to compare the incidence of UGI ulceration between celecoxib, NSAIDs and placebo. In two 12-week studies with Baseline and end-of-study endoscopies, the ulcer incidence with celecoxib at full therapeutic doses, as well as at 2-4 times the full therapeutic dose, was not significantly different from placebo but significantly lower than with naproxen. The superior GI safety of celecoxib compared to naproxen was confirmed by a 12-week serial endoscopy trial. Another 12-week serial endoscopy study demonstrated a significantly lower ulcer incidence with celecoxib compared to ibuprofen and a 6-month trial showed a significantly lower incidence of ulcers with celecoxib compared to diclofenac.

The endoscopy results were corroborated by a blinded tabulation of clinically significant UGI events (bleeding, perforation, and gastric outlet obstruction) that occurred during controlled studies. The analysis demonstrated an annual incidence of ulcer complications for celecoxib of 0.20% that was significantly less than the 1.68% seen with NSAIDs, but similar to placebo. The incidence of clinically significant UGI events for celecoxib was confirmed by an analysis of data from a large open-label safety trial.

Celecoxib was also differentiated from NSAIDs in terms of its effects on platelets, that contain only the COX-1 isoform. At doses up to 6-12 times the full therapeutic dose, celecoxib had no significant effect on platelet aggregation or bleeding time. In contrast, NSAIDs consistently and significantly inhibited platelet aggregation and prolonged bleeding time at typical therapeutic doses.

The safety of celecoxib was evaluated based on the experience with all the patients and healthy subjects who participated in clinical trials. Chronic dosing in arthritis patients ranged from 100 mg BID to 400 mg BID for periods in excess of 12 months. A dose as high as 1200 mg BID was used in certain safety/pharmacology studies. Total exposure exceeds 3,000 patient years and 981 patients have completed at least one year of treatment. Overall, celecoxib was well tolerated. GI symptoms (dyspepsia, nausea, abdominal pain) were more common in patients receiving celecoxib than in patients on placebo, but significantly less than in patients on NSAIDs. Renal adverse events were uncommon and occurred no more frequently than in patients on NSAIDs. Clinical laboratory test results indicated that celecoxib did not have adverse hematologic or hepatic effects.

Celebrex<sup>™</sup> (Celecoxib) Advisory Committee Briefing Document

In sum, celecoxib is equivalent to NSAIDs in terms of therapeutic effectiveness (a function of the inhibition of COX-2), but lacks the characteristic COX-1 dependent toxicities of NSAIDs on the GI tract and platelets. In general, celecoxib was safe and well tolerated. These results clearly establish the wide clinical therapeutic index of celecoxib and clinical utility of this new agent in the treatment of OA, RA, and pain.

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